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ASLM Conference 2014

Symposium "Innovative Solutions for HIV Diagnosis and Monitoring"

Cape Town, Wednesday 3 December 2014

Pooling of Dried Blood Spots for More Cost-Effective Viral Load Monitoring and Early Infant Diagnosis

Wolfgang Preiser, Jean Maritz,
Gert U. van Zyl, H. Newman

Division of Medical Virology
University of Stellenbosch / NHLS Tygerberg



NATIONAL HEALTH
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Fakulteit Geneeskunde en Gesondheidswetenskappe

Faculty of Medicine and Health Sciences



What is pooled testing (aka pooling) ?

Idea: Mix several individual specimens together and test the resulting "pool"

Goal: to improve affordability – use only one test for testing several samples

If test result of pool is ...

- ... negative: all individuals diagnosed as negative
- ... positive: retest all samples in pool individually to identify the positive one(s): deconvolution

Commonly used to reduce cost of screening large numbers of individuals (e.g. blood transfusion)

Considerations (I)

1. Sensitivity:

How does pooling affect test sensitivity?

We do not want to miss any (relevant) positives!

2. Pool size:

**The more samples per pool, the higher the savings;
however: The more pools test positive, the more
pools will have to be deconvoluted**

Considerations (II)

3. Prevalence:

The lower the better:

fewer individuals positive → more pools test negative → no deconvolution required

Lower prevalence → bigger pool sizes possible

4. Throughput

Considerable test volumes required for pooling to be viable; waiting to fill up runs prolongs turnaround times

Possible use: HIV viral load testing

Method of choice to monitor patients on ART



UNDETECTABLE

HOW VIRAL LOAD MONITORING
CAN IMPROVE HIV TREATMENT
IN DEVELOPING COUNTRIES



PUTTING HIV TREATMENT TO THE TEST

A PRODUCT GUIDE FOR VIRAL LOAD AND
POINT-OF-CARE CD4 DIAGNOSTIC TOOLS



World Health
Organization

GUIDELINES



CONSOLIDATED GUIDELINES ON
**THE USE OF
ANTIRETROVIRAL DRUGS
FOR TREATING AND
PREVENTING HIV INFECTION**

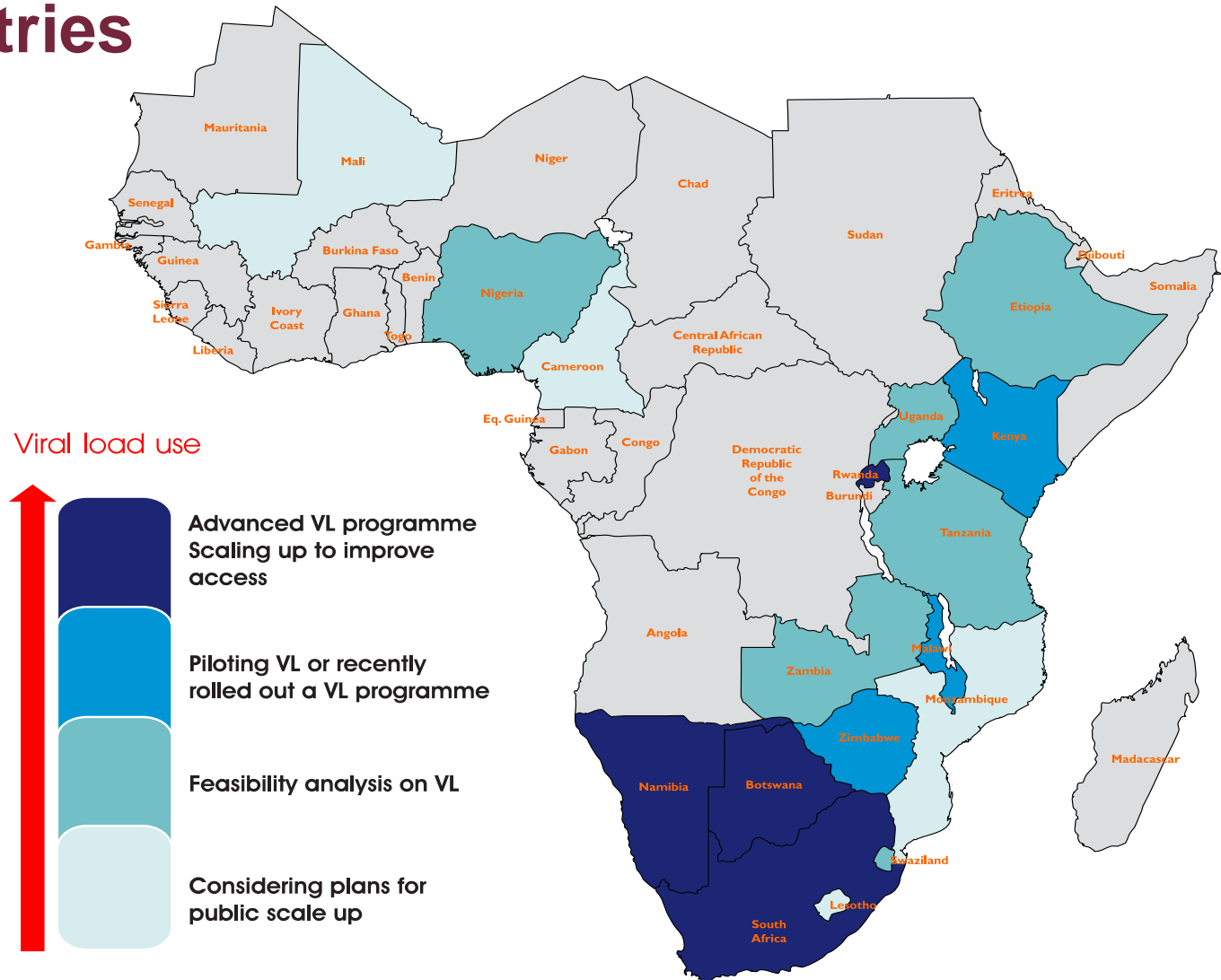
RECOMMENDATIONS FOR A PUBLIC HEALTH APPROACH

JUNE 2013

Monitoring schedule

Threshold (5000 cop./ml vs. 1000 cop./ml)

HIV VL monitoring in sub-Saharan African countries



Trevor Peter, IAS-ILF Symposium, Expanding access to viral load monitoring in resource-limited settings, Lusaka, Zambia, 2014

EID point-of-care system: Liat™ analyser (Iquum / Roche)



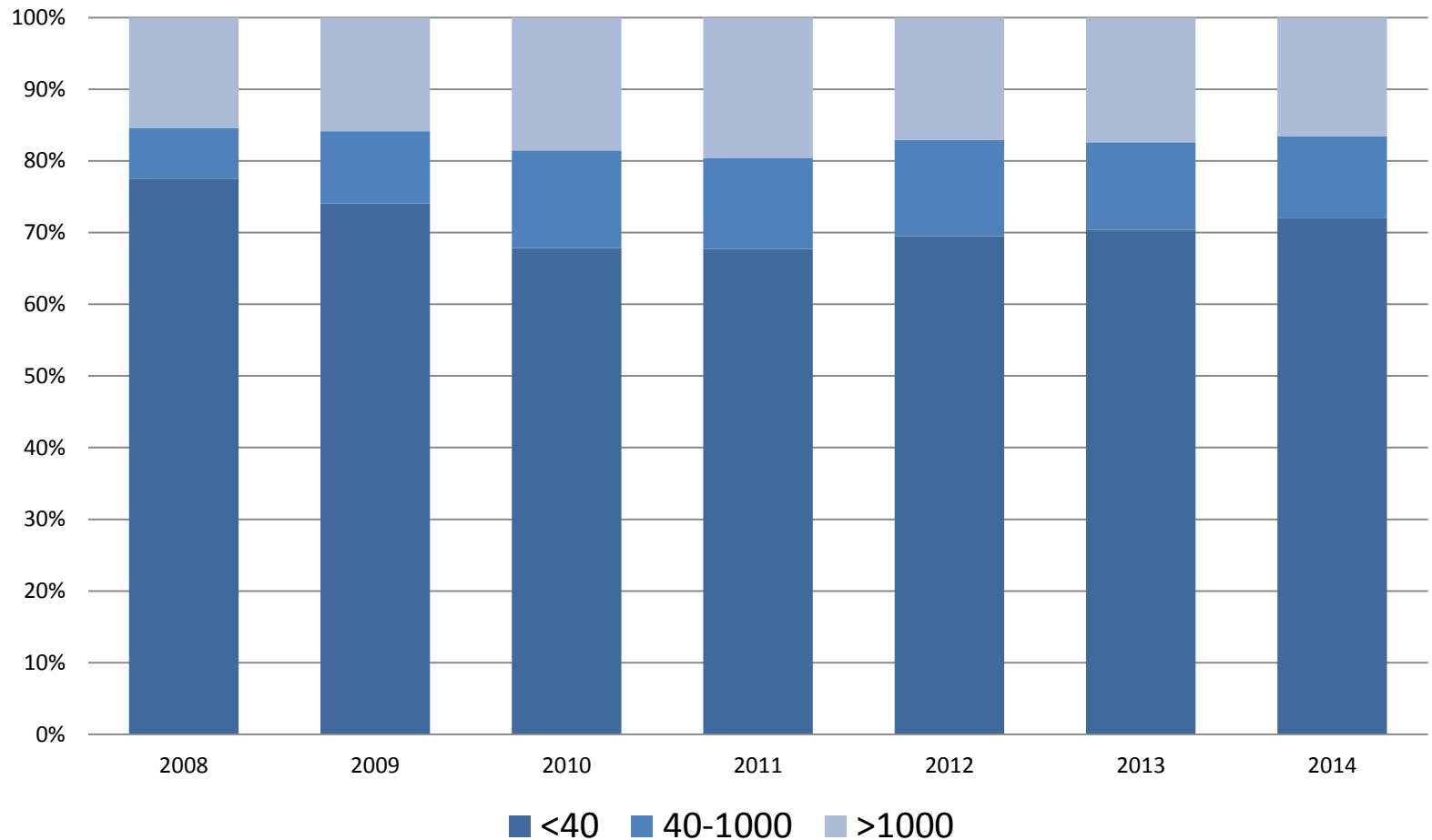


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NHLS HIV viral load platform: Abbott m2000 RealTime HIV-1 System



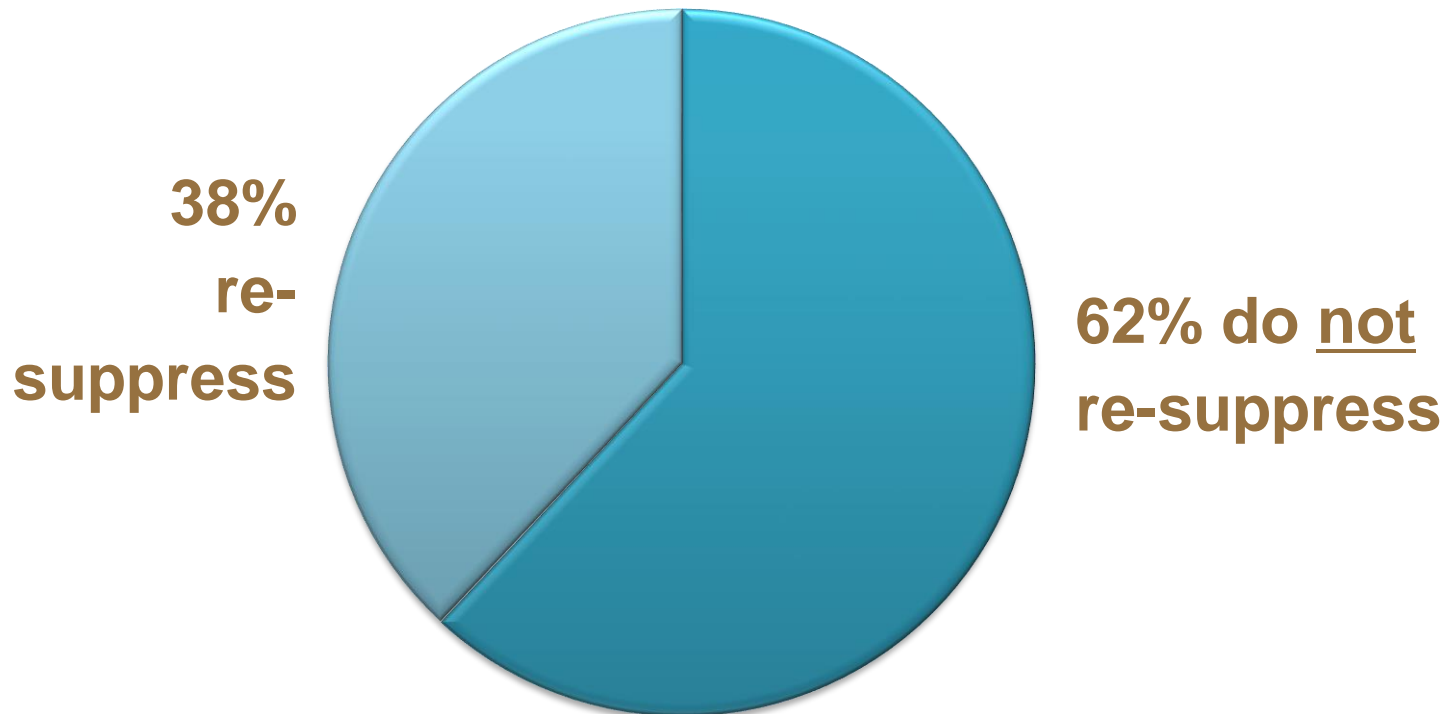
Adult HIV viral load test results, Western Cape, 2008 – 2014



Proportion of VL results >1000 copies/ml remarkably constant at just below 20%. Hsiao et al., ASLM 2014, poster 96

Adult HIV viral load test results, Western Cape, 2008 – 2014

More than 60% of samples with previous VL>1000 copies/ml failed to re-suppress, highlighting persistent ART failure as an important issue



Pooling Strategies to Reduce the Cost of HIV-1 RNA Load Monitoring in a Resource-Limited Setting

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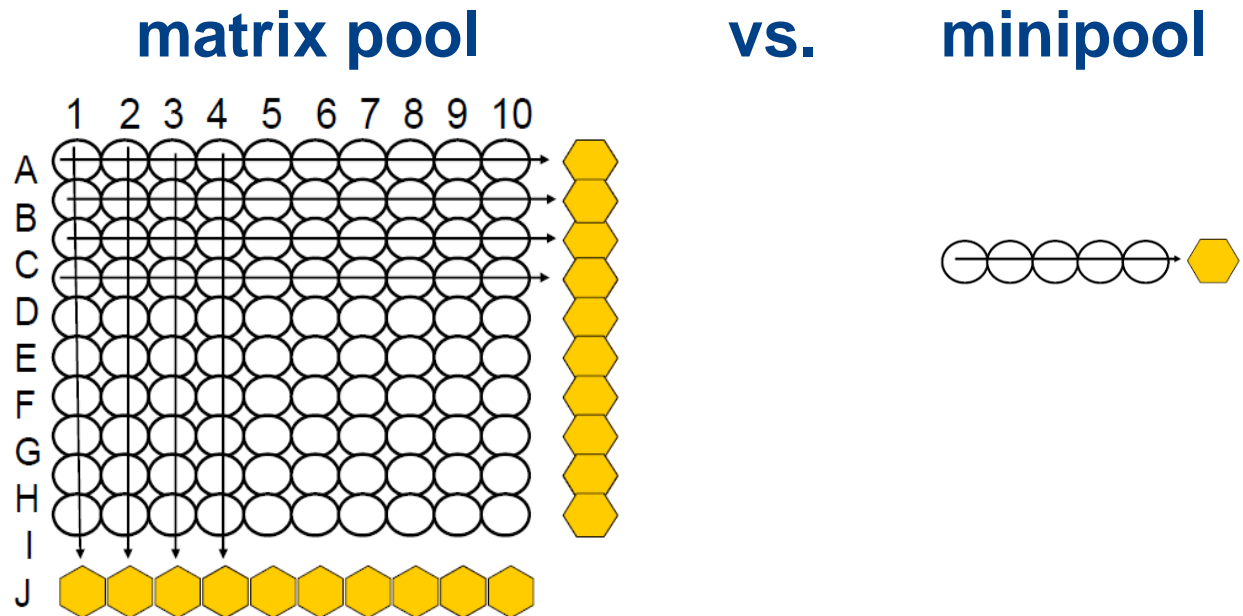
Clinical Infectious Diseases 2011;52(2):264–270

Summary: Materials & Methods

Specimens with low pre-test probability of ART failure

Plasma vs. dried blood spots vs. dried plasma spots

2 different pooling strategies:



Summary: Methods, Results

Deconvolution algorithm to identify specimens(s) with detectable viral loads

Results:

Method	% failure (>1000c/ml) /100 spec.	NPV	# tests	savings needed
3 matrices (300 specimens)	11 %	98 %	41 %	1,640 \$
80 minipools (400 specimens)	9.5 %	100 %	30.5 %	1,220 \$

Summary: Conclusions

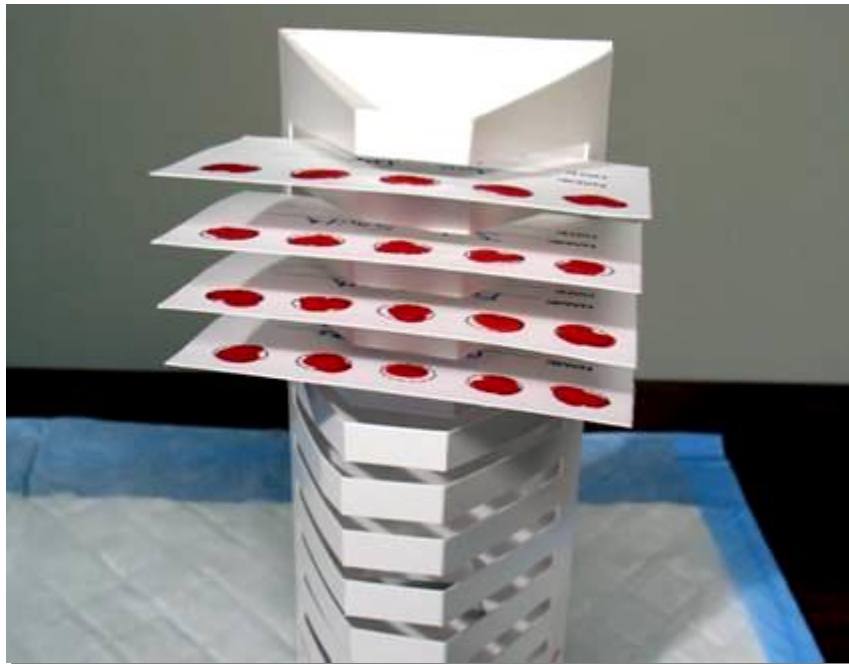
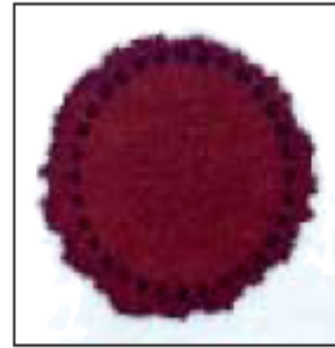
Pooling saves 30.5 % – 60 % of HIV RNA tests

Matrix strategy may be more efficient but is technically demanding

Minipools of 5 dried blood spots were accurate with NPV >95%

In resource-constrained settings, combining pre-selection of patients with low pre-test probability of virologic failure and pooled testing can reduce cost of monitoring without compromising accuracy

Suitable for dried blood spots (DBS) and dried plasma spots (DPS)

A photograph of a white card with five circular spots at the top. Below the spots are two horizontal lines for writing, labeled "NAME" and "DATE". At the bottom of the card, there is a small, faint logo or text that reads "SAS® 100% LA FINE ACTED".

Pooled HIV-1 Viral Load Testing Using Dried Blood Spots to Reduce the Cost of Monitoring Antiretroviral Treatment in a Resource-Limited Setting

Pieter Pannus, MSc, Emmanuel Fajardo, BSc,* Carol Metcalf, MBChB, MPH,*
Rebecca M. Coulborn, MPH,† Laura T. Durán, MBChB,† Helen Bygrave, MA, MBChB,*
Tom Ellman, BSc, MBChB, MSc,* Daniela Garone, MBChB, BSc (ID),† Michael Murowa, MBBS,‡
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J Acquir Immune Defic Syndr • Volume 64, Number 2, October 1, 2013

Malawi pooling study

Compared plasma vs. finger prick DBS vs. venous blood DBS

Compared "minipool" and "minipool + algorithm" strategies on pools of 5 samples

Accuracy: NPV 97.3% - 100%, PPV 96.2% - 100%

Efficiency: depends on sample type and threshold (1000 cop./ml vs. 5000 cop./ml)

Example: with finger prick DBS and 5000 cop./ml threshold, mini pooling reduces number of tests required by 51.4% compared with individual testing

And finally: Do we need the "load" in viral load?

A qualitative PCR minipool strategy to screen for virologic failure and antiretroviral drug resistance in South African patients on first-line antiretroviral therapy

Howard Newman^{a,*}, Lukas Breunig^{a,b}, Gert van Zyl^a, August Stich^b, Wolfgang Preiser^a

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Journal of Clinical Virology 60 (2014) 387–391

Pooled qualitative PCR to detect virological failure ...

**Qualitative in-house PCR targeting partial RT gene
300 routine patient samples (incl. 29 positives, of
which 26 with VL >1000 cop./ml)**

**60 minipools of 5 EDTA blood samples each
22 / 60 pools tested positive**

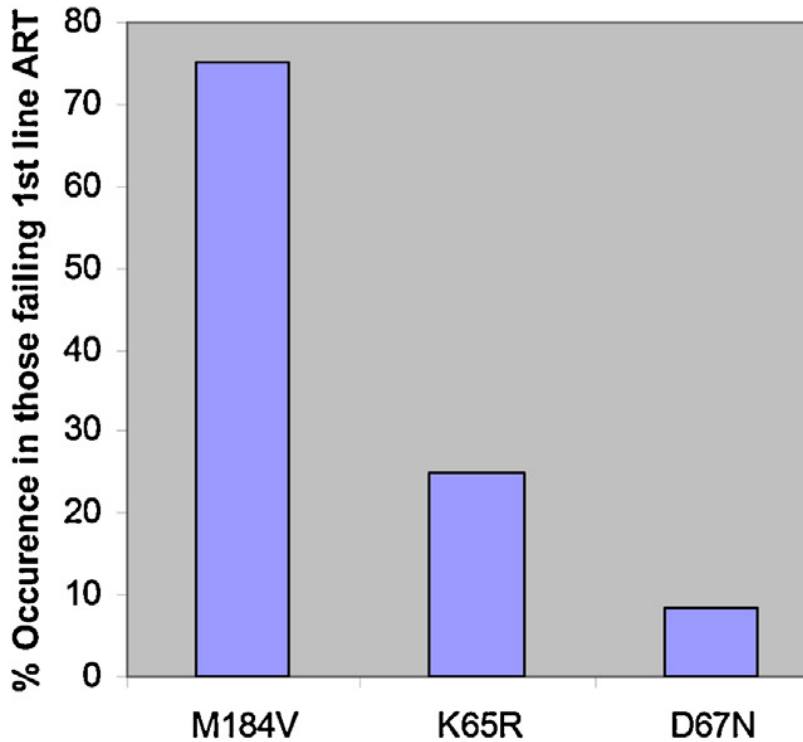
Pooling detected 24 / 26 failing patients

**Sensitivity for detecting failure 92%, specificity
98.9%**

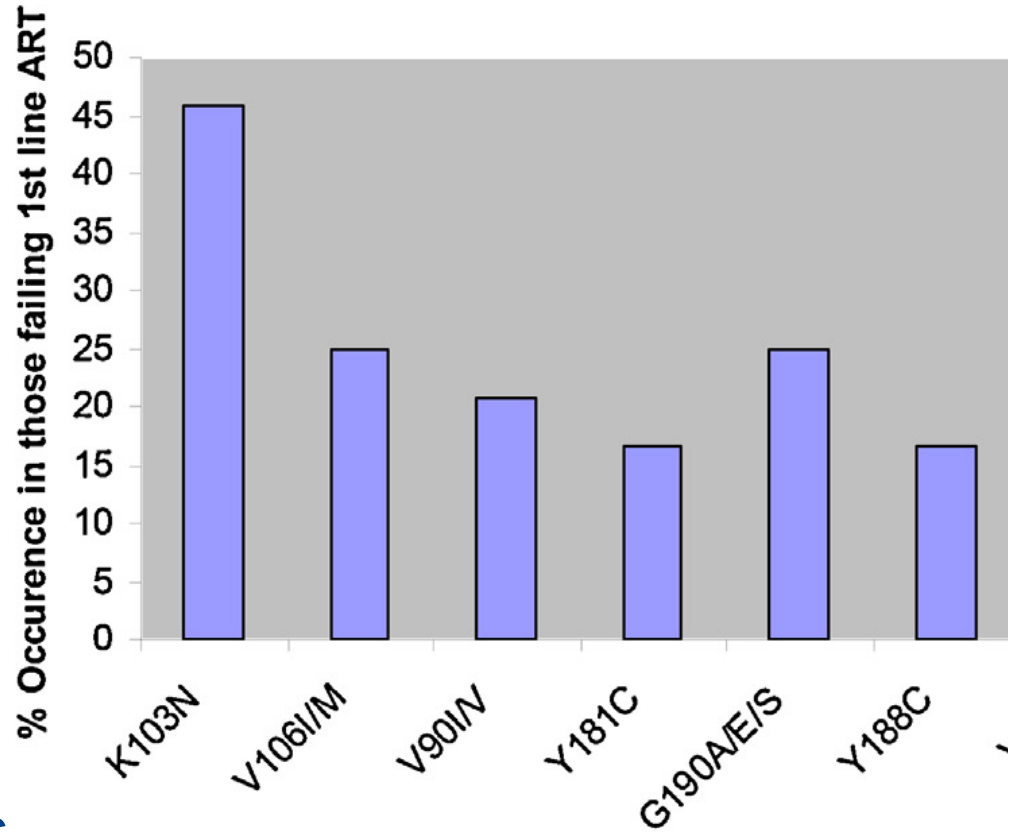
NPV 99.3%, PPV 89.7%

**Pooled testing required 43% fewer assays than
conventional viral load testing**

... and sequencing of positives



NRTI resistance mutations



NNRTI resistance mutations

Possible use: HIV early infant diagnosis (EID)

Requires detection of viral nucleic acid (proviral DNA and / or viral RNA) or viral antigen (p24) due to presence of maternal antibodies

Complex, expensive; centralised → delays

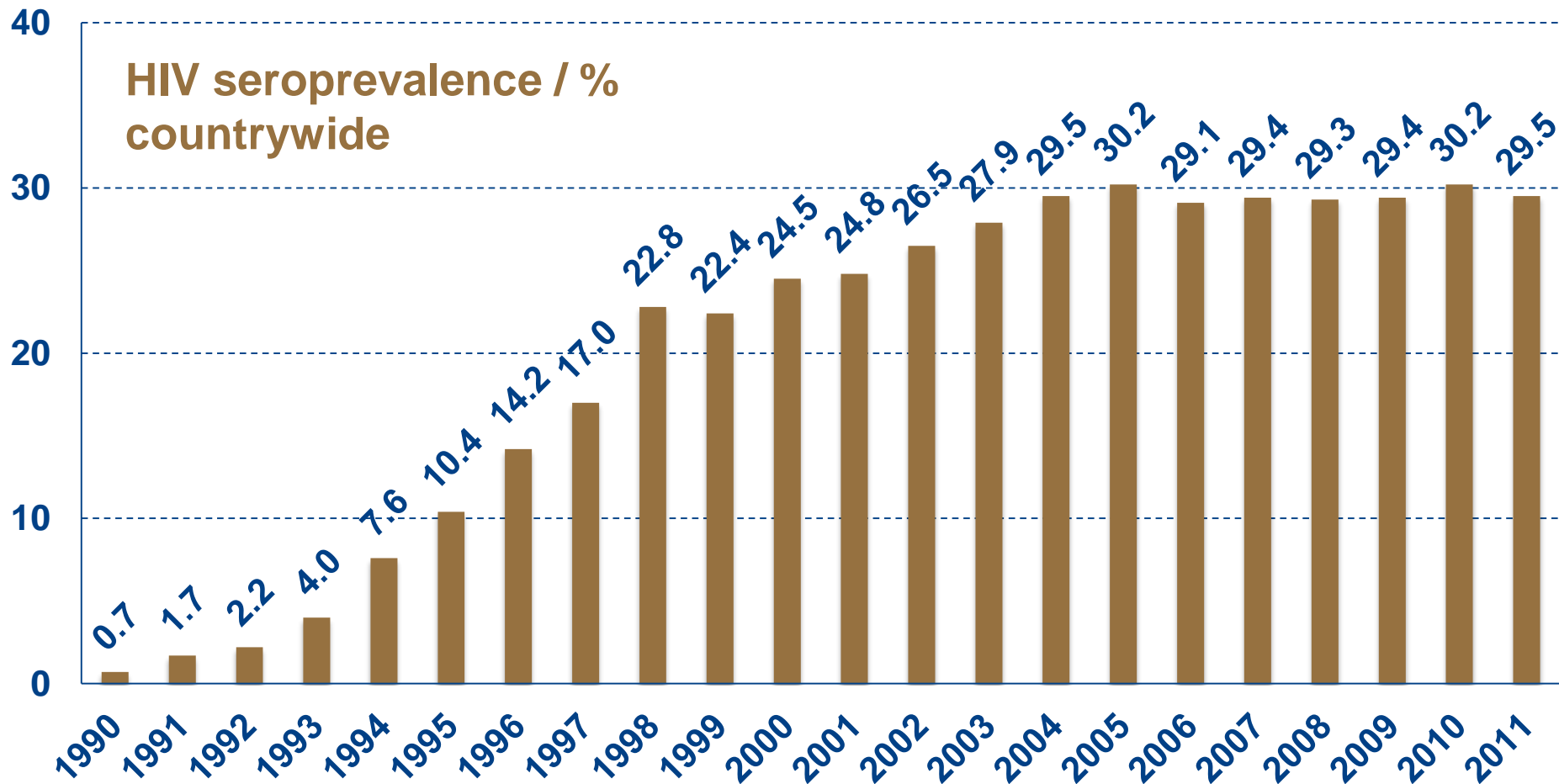
Infant testing at 6 weeks is not enough:

3/4 of HIV-infected babies can be diagnosed at birth (intra-uterine infection)

CHER study: Infant mortality peaks at 3 months of age → early ART improves outcomes!



HIV seroprevalence in pregnant women in South Africa: 1990 – 2011



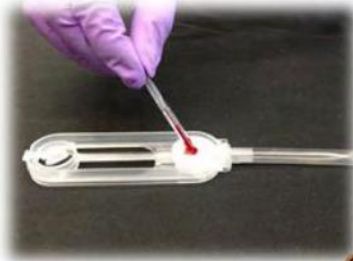
National Dept. of Health, RSA:
2011 National Antenatal Sentinel HIV & Syphilis Prevalence Survey

LYNX p24 antigen point-of-care assay (Northwestern Global Health Foundation)

Step 1: Collect blood



Step 2: Apply blood to LYNX Plasma Separator



Step 3: Plunge Plasma Collection Pad into the Reaction Tube

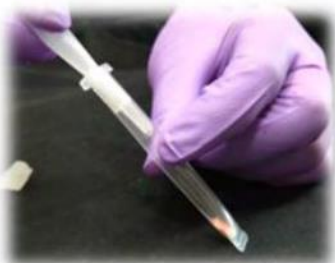


Step 4: Separate Reaction Tube from LYNX Plasma Separator



5 - 10 minutes

Step 5: Add LYNX Buffer



Step 6: Heat



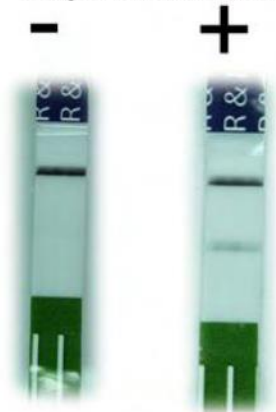
11 minutes

Step 7: Insert LYNX Test Strip



30 minutes

Step 8: Read Test



NHLS EID platform: COBAS AmpliPrep / COBAS TaqMan



Infant HIV PCR results, Western Cape, South Africa, 2008 – 2014

Year	# tests	Positive	Community positivity rate *
2008	19058	8.79%	3.55%
2009	19518	8.32%	3.54%
2010	17681	5.79%	2.86%
2011	18042	4.14%	1.52%
2012	19716	3.41%	1.36%
2013	21560	3.14%	1.24%
2014	24873*	3.26%	0.99%

* tests requested from primary level facilities for infants 5 – 7 weeks of age

Pilot study: Test results of pooled samples

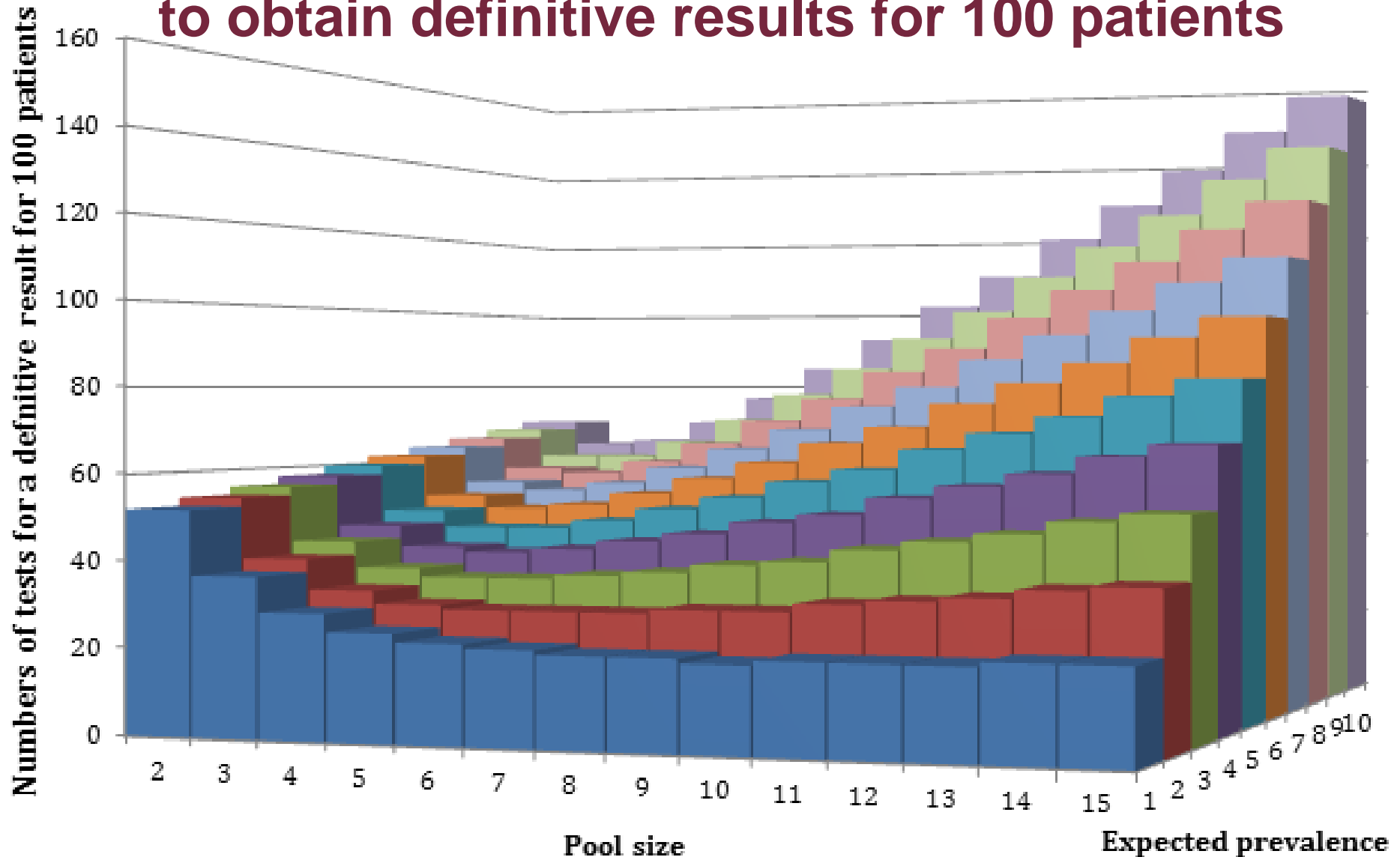
	HIV reactive pools		HIV non-reactive pools
	HIV reactive DBS	Total HIV reactive DBS	
Reactive result	35	38	0
Negative result	0	1	19
Sensitivity		97% ^γ	
Specificity			100% ^γ
Positive predictive value ^α		100 % ^γ	
Negative predictive value ^α			99.9% ^γ

^α = Calculated using Bayes' rule and a prevalence of 2.0% as determined by CAP/CTM assay tested on individual whole blood samples of 100 μL during the preceding 12-month period

^γ = Calculation based on all reactive pools, including weakly reactive DBS pools

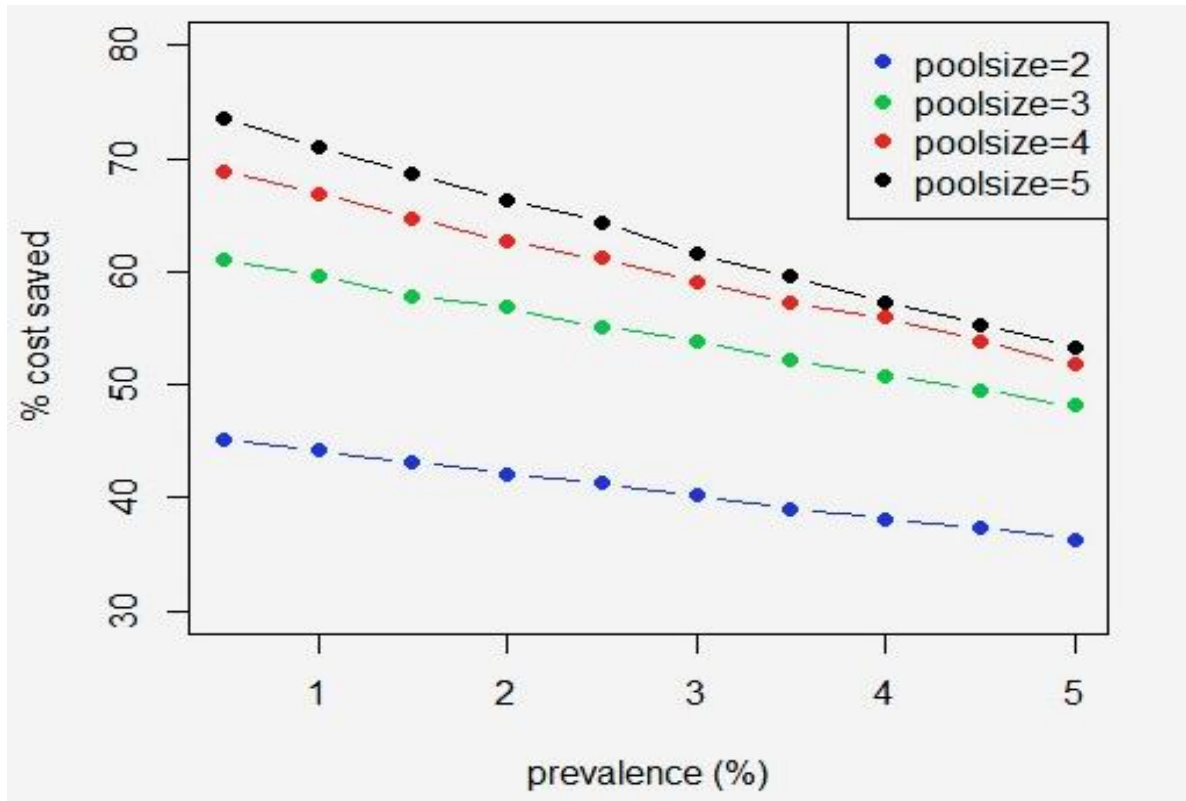
J. Maritz, S. Douma, W. Preiser, 2014 (study ongoing)

Pilot study: Maximal number of tests needed to obtain definitive results for 100 patients



Modelling

"Cost- effectiveness of pooled PCR testing of dried blood spots for infant HIV diagnosis" accepted for poster presentation at CROI2015



The next step: implementation where EID is not always available and has long TATs



Bugando Medical Centre, Mwanza, Tanzania

Conclusions: Pooled testing ...

... can help meet the enormous and largely unmet need for HIV EID and HIV viral load testing in many African settings

... is feasible if relatively few patients are infected / failing as is increasingly the case in Africa

... can be done without unacceptable decreases in sensitivity and accuracy

... reduces number of tests needed → cost reduction

Outlook

Centralised, high throughput setting: optimises use of scarce qualified staff and sophisticated facilities

Option: highly automate e.g. pipetting robot and computer-guided pooling and deconvolution

Defining appropriate pooling strategies

(pool size, pool type (mini, matrix, 3D), ...)

needs to take into account both economical and practical perspectives



Outlook

Pooled testing should be part of a "package"

Stratify patients:

- **'low risk' (e.g. those in ART adherence clubs)
→ pooled testing**
- **'high risk' (e.g. those who would receive targeted VL testing) → individual testing**

Pooled testing can improve affordability and thus availability of virological diagnosis and monitoring in resource-constrained settings

Collaborators

**Tygerberg: Jean Maritz,
Gert van Zyl, Howard
Newman, Sanne Douma**



**MSF: Pieter Pannus, Emmanuel
Fajardo, Carol Metcalf, ...**



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Davy Smith, Richard Haubrich**



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Würzburg: Gustl Stich, Lukas
Breunig, Susanne Potschka,
Anna-Theresa Lundershausen**





**Thank you, baie dankie,
enkosi kakhulu, vielen Dank!**